# EST Analysis of the Cnidarian *Acropora millepora*Reveals Extensive Gene Loss and Rapid Sequence Divergence in the Model Invertebrates

R. Daniel Kortschak, 1,2,3,5 Gabrielle Samuel, 1,5 Robert Saint,1,2,\* and David J. Miller4,\* <sup>1</sup>Centre for the Molecular Genetics of Development and Molecular Genetics and Evolution Group Research School of Biological Sciences **Australian National University** P.O. Box 475 Canberra, ACT2601 Australia <sup>2</sup>Centre for the Molecular Genetics of Development Department of Molecular Biosciences University of Adelaide Adelaide, SA5005 Australia <sup>3</sup>Centre for Bioinformation Science **Australian National University** Canberra, ACT2601 Australia <sup>4</sup>Comparative Genomics Centre Molecular Sciences Building James Cook University Townsville, Queensland 4811 Australia

# Summary

A significant proportion of mammalian genes are not represented in the genomes of Drosophila, Caenorhabditis or Saccharomyces, and many of these are assumed to have been vertebrate innovations. To test this assumption, we conducted a preliminary EST project on the anthozoan cnidarian, Acropora millepora, a basal metazoan. More than 10% of the Acropora ESTs with strong metazoan matches to the databases had clear human homologs but were not represented in the Drosophila or Caenorhabditis genomes; this category includes a surprising diversity of transcription factors and metabolic proteins that were previously assumed to be restricted to vertebrates. Consistent with higher rates of divergence in the model invertebrates, three-way comparisons show that most Acropora ESTs match human sequences much more strongly than they do any Drosophila or Caenorhabditis sequence. Gene loss has thus been much more extensive in the model invertebrate lineages than previously assumed and, as a consequence, some genes formerly thought to be vertebrate inventions must have been present in the common metazoan ancestor. The complexity of the Acropora genome is paradoxical, given that this organism contains apparently few tissue types and the simplest extant nervous system consisting of a morphologically homogeneous nerve net.

## **Results and Discussion**

Our understanding of metazoan genome evolution is based on a small number of complete genome sequences and large EST datasets that represent only a few complex animals. Of necessity, therefore, deductions about the evolutionary origins and structures of human genes are largely based on comparisons with the genomes of the insects Drosophila melanogaster and Anopheles gambiae, the nematode Caenorhabditis elegans, and the yeasts Saccharomyces cerevisiae and Schizosaccharomyces pombe. The study of the evolution of developmental genes has identified some spectacular examples of conservation of developmental programs, particularly between D. melanogaster and vertebrates. However, a significant number of D. melanogaster and C. elegans genes are highly modified, and the extent of gene loss in these organisms is unknown [1-3]. Substantial differences between the D. melanogaster and C. elegans genomes, together with the fact that the nematodes and arthropods are now generally regarded as more closely related than was previously the case [4, 5] (although see also [6]), imply that comparisons based only on these organisms may give a misleading view of the ancestral metazoan.

In terms of understanding the evolution of metazoan genetic and developmental complexity, the Cnidaria are likely to be critically important, as this phylum is regarded as the sister group to the Bilateria [7]. The limited available data suggest that cnidarian genes may not only reflect ancestral intron/exon structures [e.g., 8, 9], but also highlight gaps in our understanding of animal genome evolution. For example, our analysis of DPP pathway genes in *A. millepora* revealed a remarkable level of conservation between these genes and their vertebrate, rather than their invertebrate, counterparts [10, 11]. Jellyfish (*Podocoryne carnea*) equivalents of the myogenic genes *Brachyury*, *Mef2*, and *Snail* are also more similar to their vertebrate homologs than to their *D. melanogaster* and *C. elegans* counterparts [12].

To obtain a broader perspective on patterns of gene evolution, we conducted a limited EST analysis on the coral Acropora millepora, A. millepora is a member of the basal cnidarian class, the Anthozoa, making it an appropriate chidarian for comparative purposes [13]. A 96 hr planula larva stage cDNA library for A. millepora [14] was normalized prior to sequencing individual clones. cDNAs were amplified using vector primers, denatured at 100°C for 5 min, and then allowed to reassociate at 65°C for 24 hr prior to hydroxyapatite chromatography at 60°C in 0.12 M phosphate buffer (pH 6.8). After concentration, the eluate was used as template in PCR and products cloned into pGEM-T easy (Promega). 3024 independent clones were isolated, and 2592 single-pass sequences were obtained using the T3 primer. Using the phred/phrap package (Brent Ewing and David Gordon, University of Washington), clone sequences were filtered for vector contamination and quality. This resulted in 632 sequences being discarded. The remaining se-

<sup>\*</sup>Correspondence: david.miller@jcu.edu.au (D.J.M.), saint@rsbs.anu. edu.au (R.S.)

<sup>&</sup>lt;sup>5</sup>These authors contributed equally to this work.

quences were grouped into 1376 EST clusters using BLASTN [15]. 999 ESTs were present as unique sequences, while 961 clones formed 377 clusters of two or more clones, which were treated together in subsequent analyses. The EST sequences were compared to the GenPept database (November 7, 2002) using the BlastX program [15] with a Blast E-statistic acceptance threshold of 10<sup>-6</sup>. In total, 492 ESTs matched entries in the GenPept database with E-statistics ranging from 10<sup>-6</sup> to 10<sup>-137</sup>. The EST sequences are available at http://cbis.anu.edu.au/coral/.

## Gene Loss in Drosophila and Caenorhabditis

The most surprising implication of the Acropora dataset is that extensive gene loss has occurred in Drosophila and Caenorhabditis - a substantial number of the coral ESTs (53 clusters; 11% of hits to any organism) appeared to have a human homolog but no counterpart in the fly or worm. A much smaller number (five clusters; 1% of hits to any organism) of the sequences gave significant matches only with fly or worm genes. Table 1 lists those A. millepora ESTs with nominal homologs in only either *H. sapiens* or the model invertebrates. To assess whether these genes arose in, or predate, the metazoan lineage, the A. millepora sequences were used to search for homologous sequences in representatives of other kingdoms. Of the 58 sequences listed in Table 1, all but six lack identifiable homologs in other kingdoms, suggesting that these genes are likely to have been metazoan innovations that have subsequently been lost from at least some (metazoan) lineages.

The absence of clear D. melanogaster or C. elegans homologs of many genes represented in the coral and human genomes thus reflects secondary gene loss in the lineages that gave rise to these model invertebrates. In at least some cases, however, these genes have been assumed to be vertebrate innovations. Examples include Churchill (AmEP01258; [16]) and Tumorhead (AmEP00036; [17]), two genes involved in the regulation of early neural development in vertebrates. Such gene losses are not confined to specific functional classes, although metabolic/structural proteins and transcription factors account for most of those identified. The scale of gene loss-53 genes missing from the genomes of both fly and worm identified in this analysis of 1376 EST clusters—implies that a major fraction of nominally vertebrate-specific genes were, in fact, present in the common metazoan ancestor. Decreasing the matching stringency increases the proportion of nominal vertebrate-only matches in the Acropora dataset (data not shown; note also that there is a corresponding decrease in confidence in the Acropora/human match). Broadening the search to consider matches with other vertebrates likewise identifies additional cases of nominally vertebrate-specific genes in the coral EST dataset; for example, a clear match with snake venom phospholipase A2 is represented both in the Acropora dataset (1e<sup>-14</sup>) and in the jellyfish Cyanea [18]. No other systematic attempts have been made to assess the proportion of genes shared between cnidarians and vertebrates that are missing from Drosophila and Caenorhabditis. However, searching for specific candidates amongst the limited data available for other cnidarians allowed us to identify a number of other genes in this category; for example, homologs of the Dickkopf proteins (mostly Wnt antagonists) and p8 transcription factors can be found amongst the available Cyanea capillata ESTs [18], while a clear homolog of the human Bardet-Biedl syndrome 4 gene is present in Hydra magnipapillata (database accession number AAO72330). Nearly one-sixth of genes identified in the urochordate Ciona lack a clear Drosophila or Caenorhabditis homolog but are represented in the Fugu or human genomes [19]. Whereas these genes have been assumed exclusively to represent chordate- (or deuterostome-) specific innovations [19], the cnidarian data imply that this interpretation is probably incorrect and that although some chordatespecific genes undoubtedly exist, the number is likely to be much smaller than has been assumed to date. Consistent with this, most, but not all, of the genes listed in Table 1 as uniquely shared between Acropora and man are present in Ciona and would therefore be amongst the 2570 genes previously considered as likely to be chordate specific [19]. Hence, many genes assumed to have much later evolutionary origins are likely to have been present in the common metazoan ancestor and have been lost from the fly and worm. In some respects, this pattern of gene loss in metazoans resembles that reported for fungi; Saccharomyces cerevisiae appears to have lost about 300 genes since divergence with Schizosaccharomyces pombe [20, 21]. Although the true extent of gene loss from the model invertebrates will only become clear after comprehensive analyses of a range of non-standard animals, the complete genomic sequence of an anthozoan such as Acropora or Nematostella is likely to identify many genes previously assumed to be vertebrate innovations.

# The Derived Nature of Many Other *Drosophila* and *Caenorhabditis* Genes

A more subtle revelation of the Acropora EST dataset is the extent to which coral sequences resemble human genes rather than the corresponding Drosophila and Caenorhabditis sequences; in comparisons against the entire database, the majority of the coral ESTs show much higher similarity to vertebrate sequences than to any invertebrate sequences (Figure 1A). For clarity, we conducted comparative analyses to *H. sapiens* and the model invertebrates D. melanogaster and C. elegans, and for each, near complete genome sequences are available. Of the 455 ESTs identified with clear matches in at least one of these three metazoans, only a small proportion (33 or 7%) showed >103-fold more significant similarity to fly and/or worm sequences, as measured by BLAST E-value, while 165 (36%) showed >103-fold more significant similarity to human sequences (Figure 1B). These results confirm a trend in closer relatedness between coral and vertebrate sequences (than between coral and fly or worm sequences) that we have observed in previous studies [10, 11]. The same general trend has been observed with specific proteins from some other cnidarians (see, for example, [12]). The general trend toward higher overall similarity is not confined to specific functional classes of genes. It is true of genes that encode

Table 1. A. millepora ESTs, Which in Comparisons with H. sapiens, D. melanogaster, and C. elegans, Are Uniquely Shared with Either Man Or the Model Invertebrates

AmEP00013	Similar to RIKEN 1700003M02 (unknown function)	BC035083	6e-19
AmEP00017	Uncharacterized WD, SAM and U box domain protein	BC029520	2e-16
AmEP00036	Tumorhead (Xenopus) homolog	AK055726	6e-8
AmEP00100	Cystatin B (stefin B)	BC010532	2e-21
AmEP00144	MBT-related	AL110249	8e-8
AmEP00176	Unknown function (contains OST3/OST6 domain)	AF161425	7e-16
AmEP00191	t-sec1 (mouse) homolog	BC028119	3e-12
AmEP00195	L-arginine:glycine amidinotransferase	X86401	6e-57
AmEP00227	Agmatinase (agmatine ureohydrolase)	AY057097	1e-44
AmEP00233	Unknown function	AK098840	7e-21
AmEP00235	M phase phosphoprotein 6	BC031017	4e-20
AmEP00265	Alpha 1 type III collagen	BC028178	5e-18
AmEP00274	Cytochrome c oxidase subunit VIIc	BC007498	2e-8
AmEP00280	Dolichyl-phosphate mannosyltransferase polypeptide 3	BC032223	8e-13
AmEP00301	Retinol dehydrogenase 8 (all-trans)	AK024022	1e-6
AmEP00320	Chromosome 22 ORF 23 UPF0193/Pfam05250/EVG1	BC031998	2e-8
AmEP00322	Alpha X integrin (Cd11c/leukocyte adhesion p150)	Y00093	7e-7
AmEP00346	PCAF associated factor 65 alpha (TAF6L)	AF069735	6e-15
AmEP00352	Unknown function	BC031107	2e-11
AmEP00422	Hypothetical protein FLJ90575 (unknown function)	AK075056	3e-32
AmEP00443	Subtilisin/kexin isozyme SKI1-related	AL133583	1e-7
AmEP00443	Zinc finger protein 294	AB018257	4e-7
AmEP00451	HMG protein HMGX1	AF146222	6e-18
AmEP00510	PRTD-NY2 (contains RGS domain)	AY009106	2e-11
AmEP00564	HGF activator precursor	D14012	4e-8
AmEP00589	Microtubule actin crosslinking factor 1 isoform 4	AB007934	2e-12
AmEP00622	KPL2 (rat)-related	AK027230	4e-12
AmEP00640	Adenylate kinase 5-related	AK027230 AK090967	2e-8
AmEP00640 AmEP00671	KNP-I/ES1 (zebrafish) homolog	BC003587	4e-24
AmEP0077	N-methylpurine DNA glycosylase	AF499437	4e-24 4e-31
AmEP00759	Glutathione S-transferase Mu4	X68677	4e-16
AmEP00762	Zinc finger protein 291	AF242528	1e-19
AmEP00772	Microsomal glutathione S-transferase 3	BC005964	6e-29
AmEP00781	Methyl-CpG binding protein 2	BC032638	4e-20
AmEP00808	Cysteine rich FGF receptor	U28811	1e-7
AmEP00831	Unknown function	AK094715	4e-21
AmEP00843	Similar to Ts mitochondrial translation EF	BC022862	5e-11
AmEP00909	NYD-SP14 (unknown function)	BC024193	9e-13
AmEP00926	Hypothetical protein FLJ21977 (ELMO-related)	BC010991	8e-10
AmEP00972	Chromosome 4 ORF45	AL834487	2e-18
AmEP00986	Acid phosphatase type 5	J04430	6e-10
AmEP00998	Unknown function	AK092816	4e-18
AmEP01004	Similar to KIAA0543 protein (unknown function)	AC004877	5e-7
AmEP01027	Neuronal pentraxin 1	U61849	2e-9
AmEP01064	Serine dehydratase-related	BC009849	2e-15
AmEP01203	Inositol polyphosphate 4-phosphatase	U26398	7e-7
AmEP01241	Unknown function	BC004228	8e-15
AmEP01258	Churchill (mouse) homolog	BC020550	1e-27
AmEP01267	Spasmolytic polypeptide (trefoil domain containing)	X51698	5e-14
AmEP01274	Related to retinitis pigmentosa 9	BC025928	7e-26
AmEP01283	Kinectin 1	Z22551	4e-11
AmEP01304	Thymosin beta-10	M92383	1e-10
AmEP01317	Unknown function (possible Smc domain)	AB040938	2e-22
	th Drosophila or Caenorhabditis:	AB040300	26-22
AmEP00030	Drosophila CG3698 (unknown function)	NM_141000	9e-15
AmEP00080	Drosophila CG10933 (unknown function; SH3 domains)	NM_137407	1e-12
AmEP00080 AmEP00223	Drosophila Pao retrotransposon peptidase	NM_137407 U23420	1e-12 4e-9
AmEP00223	Caenorhabditis (putative secreted or extracellular)		
AmEP01217	Drosophila CG7102 (unknown function; TLDc domain)	NM_077448 AY118856	1e-7 3e-9
, C I C I Z I I	2.000pima Odi 102 (dimilowii idilolioli, 1200 dollidili)	A1110000	00 0

The left column gives the *A. millepora* EST cluster number. The best match on BlastX comparisons with the database are given, along with the corresponding Genbank database identifier and the e value. To confirm that those sequences identified are lineage restricted, secondary database comparisons were carried out. Human (or fly or worm) proteins matching the coral ESTs were themselves used to search the databases (using BlastP) followed by manual inspection of the results.

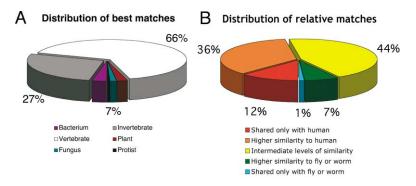


Figure 1. Summary of *Acropora* EST Sequence Comparisons

(A) Distribution of ESTs giving a best match to specific organism classes. In this case, the proportions of the 492 EST clusters giving strong database matches are indicated.

(B) Distribution of relative matches amongst the representative invertebrates (*D. melanogaster* and *C. elegans*) and *H sapiens*. In this case, the dataset consisted of the 397 ESTs with both fly/worm and human matches and the 58 ESTs that had only either fly/worm or human matches (see Table 1 for a breakdown of the latter category). Note that the phrase ">Higher similarity..." refers to >10³-fold

lower BLAST-E values (see text). Although a significant number of matches were with bacterial sequences, these are unlikely to represent contamination as the library was generated from polyA<sup>+</sup> mRNA [14], and the best matches were with taxonomically diverse bacteria.

transcription factors, such as DC6/E(y)2 (AmEP01369), Hex (AmEP00044), Y box transcription factor (AmEP00530) and Snail (AmEP00427), as well as for components of cellular signaling pathways, such as DPP, Smads and Delta (AmEP00563), and for housekeeping proteins such as methyl CpG binding protein (AmEP00781) and WASp (AmEP00290). In many cases, the highest levels of identity were between coral and human sequences, with the worm and fly typically showing higher levels of divergence. Figure 2 shows phylogenetic analyses of some representative coral sequences, illustrating the general trend of much higher levels of identity with human than with fly or worm sequences.

Although a close relationship between coral and human sequences is superficially surprising given that the cnidarian and bilaterian lineages are thought to have diverged in deep Precambrian time [22], this is largely a consequence of the relative pace of change in the model invertebrates. The greater divergence in D. melanogaster and C. elegans sequences is unlikely to reflect uniform rates of change over long time periods; rather, rapid genome change is likely to have occurred recently (and probably independently) in these organisms and be associated with intense selection for small genome size, rapid developmental rates, and the highly specialized lifestyles of the fly and worm. Although D. melanogaster had the previously reported fastest rate of sequence change, the genes of C. elegans are evolving even faster [23, 24], and genome rearrangements are occurring approximately four times faster in the worm than in the fly [24]. Typically long branch lengths in phylogenetic analyses (see Figure 2) support the idea that many D. melanogaster sequences are highly derived relative to their coral and human counterparts. This is also true of many C. elegans sequences [23, 25]. If this hypothesis is correct, we might expect sequences from more "primitive," and so less-derived, protostomes to be more closely related to the coral/human gene set; Hox data for the ribbonworm Lineus are consistent with this hypothesis [26]. Comparisons with lophotrochozoans and with the cephalochordate amphioxus will be particularly informative; however, at present these are poorly represented in the databases. Comparisons with the urochordate Ciona [19] emphasize the derived position of the model invertebrates and, although only limited comparative data are available for representatives of the more basal insect orders, these often also dramatically support the derived position of *D. melanogaster*. For example, although *D. melanogaster* does not carry out standard CpG methylation and lacks typical MBD proteins [27], more primitive insects such as the cricket *Acheta* are more vertebrate-like in both respects (see Figure 2C; [28]). Similarly, comparisons of retinoic acid receptor ligand binding domains (RXR LBDs) indicate that tick, crab, and grasshopper (*Locusta migratoria*) sequences are more similar to their vertebrate orthologs than to the *D. melanogaster* LBD or those of other holometabolous insects [29].

# **Genetic Complexity and the Common Ancestor**

A third implication of our analyses is that at least at the level of gene complement, the ancestral metazoan is likely to have been much more complex than was previously imagined. For example, the A. millepora EST dataset contains homologs of many bilaterian genes whose specialized functions are associated with highly differentiated nervous systems. These include genes with vertebrate, but no known invertebrate, counterparts; e.g., those that encode photoreceptor all-transretinol dehydrogenase (AmEP00301), Churchill, and Tumorhead. They also include more generally conserved homologs of genes that encode Frequenin, Homer 2d, Glia maturation factor b, and Notch pathway components. This complexity is particularly surprising given the morphological simplicity of the coral nervous system (anthozoans have the simplest extant nervous systemsmorphologically homogeneous nerve nets) and the absence of recognizable photoreceptors. Nevertheless, coral larvae display phototactic behavior [30] and the Pax-6-related gene PaxC is expressed in a subset of A. millepora presumed neurons [31]. The detection of ESTs matching hex and snail, genes that play key roles in endoderm and mesoderm patterning in triploblastic animals, supports the renewed interest in the nature of the cnidarian primary tissue layers [12]. At the very least, these findings provide a strong argument for developing a much better understanding of cnidarian developmental mechanisms, if we are to understand the origins of these mechanisms.

# **Conclusions**

Our preliminary survey of the expressed sequences of planula stage *Acropora millepora* appears to turn upside

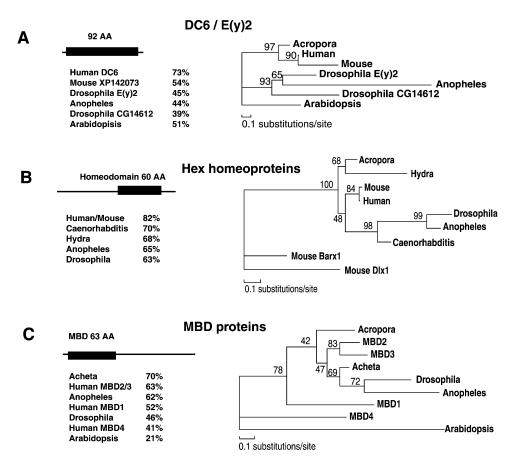


Figure 2. Comparison of Coral Sequences Indicates Higher Levels of Identity with Human Than with *D. melanogaster* or *C. elegans* Sequences Maximum likelihood phylogenetic analyses of the conserved protein domains were conducted in MolPhy version 2.3 (Institute of Statistical Mathematics, Tokyo) [32] using the Dayhoff substitution matrix and local rearrangement search mode. In each case, the percent identity with the *A. millepora* sequence is indicated below the protein schematic, and the figures on branches in the phylogenetic trees indicate percent of 1000 (ML) bootstrap replicates supporting the topology shown.

(A) A. millepora EST (AmEP01369) shows high levels of identity with human and mouse DC6 (dendritic cell protein 6), whereas the D. melanogaster enhancer of yellow [E(y)2] protein has lower identity with both the A. millepora and vertebrate sequences. C. elegans does not appear to have a member of this orthologous group; the most closely related sequence is a hypothetical protein H05L14.2/Z99772/Z75533, which has only a partial match.

(B) The *A. millepora* Hex homeodomain has higher levels of identity with its vertebrate rather than with its *Hydra* homologs. However, in ML phylogenetic analyses, the cnidarian sequences form a monophyletic group, showing that the hydra sequence is highly derived.

(C) A. millepora EST AmEP00781 encodes a typical methyl-CpG binding domain most closely related to vertebrate MBD1 and MBD2. The corresponding D. melanogaster protein (dMBD2/3) is highly derived, and no related protein has been identified in C. elegans. Human MBD4 is an atypical member of this family that functions in DNA repair rather than chromatin silencing.

down several preconceived ideas about the evolution of animal genomes. Rather than being simple, the common metazoan ancestor was genetically complex, containing many genes previously considered to be vertebrate innovations. In addition, *Acropora* and human sequences are often surprisingly similar. These data are a provocative reminder of the limited extent of our understanding of metazoan genome evolution and the potential hazards associated with extrapolating general evolutionary principles based on the model invertebrates. Whereas gene losses and modifications may obscure the picture in the model organisms and much of the animal kingdom remains to be explored, *Acropora millepora* provides a unique insight into the unexpectedly deep evolutionary origins of at least some vertebrate gene families.

# Acknowledgments

We thank Eldon Ball for commenting on the manuscript and the Australian Research Council and Australian Genome Research Facility for support. G.S. was supported by an Overseas Postgraduate Research Award.

Received: August 27, 2003 Revised: October 21, 2003 Accepted: October 21, 2003 Published: December 16, 2003

# References

- Blaxter, M. (1998). Caenorhabditis elegans is a nematode. Science 282, 2041–2046.
- 2. Ruvkun, G., and Hobert, O. (1998). The taxonomy of develop-

- mental control in Caenorhabditis elegans. Science 282, 2033–2041.
- Zdobnov, E.M., von Mering, C., Letunic, I., Torrents, D., Suyama, M., Copley, R.R., Christophides, G.K., Thomasova, D., Holt, R.A., Subramanian, G.M., et al. (2002). Comparative genome and proteome analysis of *Anopheles gambiae* and *Drosophila melanogaster*. Science 298, 149–159.
- Aguinaldo, A.M., Turbeville, J.M., Linford, L.S., Rivera, M.C., Garey, J.R., Raff, R.A., and Lake, J.A. (1997). Evidence for a clade of nematodes, arthropods and other moulting animals. Nature 387, 489–493.
- de Rosa, R., Grenier, J.K., Andreeva, T., Cook, C.E., Adoutte, A., Akam, M., Carroll, S.B., and Balavoine, G. (1999). Hox genes in brachiopods and priapulids and protostome evolution. Nature 399. 772–776.
- Blair, J.E., Ikeo, K., Gojobori, T., and Hedges, S.B. (2002). The evolutionary position of nematodes. BMC Evol. Biol. 2, 7.
- Holland, P.W. (1999). The future of evolutionary developmental biology. Nature (London) 402 (6761 Suppl), C41–C44.
- Schmitt, D.M., and Brower, D.L. (2001). Intron dynamics and the evolution of integrin beta-subunit genes: maintenance of an ancestral gene structure in the coral, *Acropora millepora*. J. Mol. Evol. 53, 703–710.
- Spafford, J.D., Spencer, A.N., and Gallin, W.J. (1999). Genomic organization of a voltage-gated Na+ channel in a hydrozoan jellyfish: insights into the evolution of voltage-gated Na+ channel genes. Receptors Channels 6, 493–506.
- Samuel, G., Miller, D.J., and Saint, R. (2001). Conservation of a DPP/BMP signaling pathway in the nonbilateral cnidarian Acropora millepora. Evol. Dev. 3, 241–250.
- Hayward, D.C., Samuel, G., Pontynen, P.C., Catmull, J., Saint, R., Miller, D.J., and Ball, E.E. (2002). Localized expression of a dpp/BMP2/4 ortholog in a coral embryo. Proc. Natl. Acad. Sci. USA 99, 8106–8111.
- Spring, J., Yanze, N., Josch, C., Middel, A.M., Winninger, B., and Schmid, V. (2002). Conservation of Brachyury, Mef2, and Snail in the myogenic lineage of jellyfish: a connection to the mesoderm of bilateria. Dev. Biol. 244, 372–384.
- Miller, D.J., and Ball, E.E. (2000). The coral Acropora: what it can contribute to our knowledge of metazoan evolution and the evolution of developmental processes. Bioessays 22, 291–296.
- Brower, D.L., Brower, S.M., Hayward, D.C., and Ball, E.E. (1997).
   Molecular evolution of integrins: genes encoding integrin beta subunits from a coral and a sponge. Proc. Natl. Acad. Sci. USA 19, 9182–9187.
- Altschul, S.F., Madden, T.L., Schaffer, A.A., Zhang, J., Zhang, Z., Miller, W., and Lipman, D.J. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res. 25, 3389–3402.
- Sheng, G., Dos Reis, M., and Stern, C.D. (2003). Churchill, a zinc finger transcriptional activator, regulates the transition between gastrulation and neurulation. Cell 115, 603–614.
- Wu, C.F., Nakamura, H., Chan, A.P., Zhou, Y.H., Cao, T., Kuang, J., Gong, S.G., He, G., and Etkin, L.D. (2001). Tumorhead, a Xenopus gene product that inhibits neural differentiation through regulation of proliferation. Development 128, 3381– 3393.
- Yang, Y., Cun, S., Xie, X., Lin, J., Wei, J., Yang, W., Mou, C., Yu, C., Ye, L., Lu, Y., et al. (2003). EST analysis of gene expression in the tentacle of Cyanea capillata. FEBS Lett. 538, 183–191.
- Dehal, P., Satou, Y., Campbell, R.K., Chapman, J., Degnan, B., De Tomaso, A., Davidson, B., Di Gregorio, A., Gelpke, M., Goodstein, D.M., et al. (2002). The draft genome of Ciona intestinalis: insights into chordate and vertebrate origins. Science 298, 2157–2167.
- Aravind, L., Watanabe, H., Lipman, D.J., and Koonin, E.V. (2000). Lineage-specific loss and divergence of functionally linked genes in eukaryotes. Proc. Natl. Acad. Sci. USA 97, 11319– 11324.
- Braun, E.L., Halpern, A.L., Nelson, M.A., and Natvig, D.O. (2000).
   Large-scale comparison of fungal sequence information: mechanisms of innovation in *Neurospora crassa* and gene loss in *Saccharomyces cerevisiae*. Genome Res. 10, 416–430.

- Conway-Morris, S. (2000). The Cambrian "explosion": slow-fuse or megatonnage? Proc. Natl. Acad. Sci. USA 97, 4426–4429.
- Mushegian, A.R., Garey, J.R., Martin, J., and Liu, L.X. (1998). Large-scale taxonomic profiling of eukaryotic model organisms: a comparison of orthologous proteins encoded by the human, fly, nematode, and yeast genomes. Genome Res. 8, 590–598.
- Coghlan, A., and Wolfe, K.H. (2002). Fourfold faster rate of genome rearrangement in nematodes than in *Drosophila*. Genome Res. 12. 857–867.
- Ledent, V., and Vervoort, M. (2001). The basic helix-loop-helix protein family: comparative genomics and phylogenetic analysis. Genome Res. 11. 754–770.
- Kmita-Cunisse, M., Loosli, F., Bierne, J., and Gehring, W.J. (1998). Homeobox genes in the ribbonworm *Lineus sanguineus*: evolutionary implications. Proc. Natl. Acad. Sci. USA 95, 3030–3035.
- Roder, K., Hung, M.-S., Lee, T.-L., Lin, T.-Y., Xiao, H., Isobe, K.-I., Juang, J.-L., and Shen, C.-K.J. (2000). Transcriptional repression by *Drosophila* methyl-CpG-binding proteins. Mol. Cell. Biol. 20, 7401–7409.
- Tweedie, S., Ng, H.-H., Barlow, A.L., Turner, B.M., Hendrich, B., and Bird, A. (1999). Vestiges of a DNA methylation system in Drosophila melanogaster? Nat. Genet. 23, 389–390.
- Hayward, D.C., Bastiani, M.J., Trueman, J.W., Truman, J.W., Riddiford, L.M., and Ball, E.E. (1999). The sequence of *Locusta* RXR, homologous to *Drosophila* Ultraspiracle, and its evolutionary implications. Dev. Genes Evol. 209, 564–571.
- Harrison, P.L., and Wallace, C.C. (1990). Reproduction, recruitment and dispersal of scleractinian corals. In Ecosystems of the World, Volume 25, Coral Reefs, Z. Dubinsky, ed. (Amsterdam: Elsevier), pp. 133–207.
- Miller, D.J., Hayward, D.C., Reece-Hoyes, J., Scholten, I., Catmull, J., Gehring, W.J., Larsen, J.E., and Ball, E.E. (2000). Pax gene diversity in the basal cnidarian *Acropora millepora* (Cnidaria, Anthozoa): implications for the evolution of the Pax gene family. Proc. Natl. Acad. Sci. USA 97, 4475–4480.
- Adachi, J., and Hagesawa, M. (1996). MOLPHY version 2.3: program for molecular phylogenetics based on maximum likelihood. Comput. Sci. Monogr. 28, 1–150.